NAGY, J.I., LEE, T., SEEMAN, P. & FIBIGER, H.C. (1978).
Direct evidence for presynaptic and postsynaptic dopamine receptors in brain. *Nature Lond.* 274, 278–281.

TITELER, M., WEINREICH, P., & SEEMAN, P. (1977). New detection of brain dopamine receptors with [³H]dihydroergocriptine. *Proc. Nat. Acad. Sci. USA.* 74/9, 3750–3753.

Binding of [3 H]-dihydroergocryptine to α -adrenoceptors on intact human platelets

D.J. BOULLIN & J.M. ELLIOTT

MRC Unit and University Department of Clinical Pharmacology, Radcliffe Infirmary, Oxford OX2 6HE

[³H]-Dihydroergocryptine ([³H]-DHE) has been shown to bind to α-adrenoceptors in both nervous (Greenberg & Snyder, 1977) and non-nervous (Williams, Mullikin & Lefkowitz, 1976; Guellaen, Yates-Aggerbeck, Vauquelin, Strosberg & Hanoue, 1978) tissue. The existence of α-adrenoceptors on the human platelet has been shown indirectly by the actions of adrenaline which induces platelet aggregation (O'Brien, 1963; Mills & Roberts, 1967) and inhibits PGE₁-induced cAMP production (Marquis, Becker & Vigdahl, 1970; Jakobs, Saur & Schultz, 1976) *in vitro*. This communication describes the direct binding of [³H]-DHE to α-adrenoceptors on intact human platelets.

Platelet rich plasma from male volunteers (age 24-38) was prepared by centrifugation at 20°C. This was centrifuged at 1700 g for 5 min at 10°C to produce a platelet pellet, which was gently resuspended in incubation buffer (0.1% EDTA, 150 mm NaCl, pH 7.5) to a final cell density of approximately 0.8×10^8 platelets/ml. One ml aliquots were incubated for 20 min at 37°C with 0.5-18 nm [3H]-DHE. Incubations were terminated by centrifugation at 1200 g for 1 min in an Eppendorf 5412 centrifuge to separate cells and supernatant. Individual platelet pellets were washed and sonicated in 500 µl distilled water; 400 µl aliquots were counted in a liquid scintillation spectrometer for total radioactivity. Specific binding was calculated as the total radioactivity bound at a given free concentration minus the non-specific binding which occurred at that free concentration in the presence of phentolamine (5 µm) (see below).

Specific binding of [3H]-DHE has an exponential onset $(T_{\frac{1}{2}} = 7.5 \text{ min for } 2.4 \text{ nm DHE})$ and offset $(T_{\frac{1}{2}} = 16 \text{ min})$. If binding were of the simple Langmuir type these values would imply that the rate constants for association (k_1) and dissociation (k_2) were $3.2 \times 10^5 \text{ m}^{-1} \text{ s}^{-1}$ and $0.74 \times 10^{-3} \text{ s}^{-1}$. The equilibrium binding curve showed a simple hyperbolic (Langmuir) form (Scatchard analysis, 5 subjects) with an affinity constant $K_a = 3.48 \times 10^8 \text{ m}^{-1}$ and binding capacity $C = 72 \text{ fmol}/10^8 \text{ platelets}$. This is equivalent to 433

molecular binding sites per platelet. The binding estimate of K_a agrees closely with the value derived from the kinetic measurements ($K_a = k_1/k_2 = 4.34 \times$ 108 m⁻¹). [3H]-DHE binding in a single subject showed little variability during a five week period, $K_a = 3.046$ $\pm 0.170 \times 10^{8} M^{-1}$ (mean \pm s.e. mean), capacity = 70.31 ± 5.15 f mol 10^{-8} platelets (mean \pm s.e. mean, n = 5). [3H]-DHE is inhibited stereospecifically by adrenaline and noradrenaline, with (-)isomer being at least ten times more potent than the (+)isomer. In comparison isoprenaline, dopamine and serotonin are poor inhibitors. Phentolamine and yohimbine at high concentrations (greater than 10⁻⁵m) appear to displace non-specifically bound [3H]-DHE. Between 10⁻⁶M and 10⁻⁵M phentolamine [³H]-DHE binding is constant, therefore a concentration of 5 µm phentolamine was used to define non-specific binding.

These results resemble those obtained on platelet lysates (Newman, Williams, Bishopric & Lefkowitz, 1978; Alexander, Cooper & Handin, 1978) except that the dose-dependent effects of phentolamine have not been reported.

We conclude that the binding of [3 H]-DHE on intact human platelets occurs at α -adrenoceptor sites and that this technique may be used to monitor the platelet α -adrenoceptor characteristics of subjects in a variety of clinical conditions.

References

ALEXANDER, R.W., COOPER, B. & HANDIN, R.I. (1978). Characterization of the human platelet α-adrenergic receptor. J. Clin. Invest. 61, 1136-1144.

GREENBERG, D.A. & SNYDER, S.H. (1977). Selective labelling of α-noradrenergic receptors in rat brain with [³H]-dihdryoergokryptine. *Life*. *Sci.* **20**, 927–932.

GUELLAEN, G., YATES-AGGERBECK, M., VAUQUELIN, G., STROSBERG, D. & HANOUE, J. (1978). Characterization with [³H]-dihydroergocryptine of the α-adrenergic receptor of the hepatic plasma membrane. *J. Biol. Chem.*, **253**, 114–1120.

JAKOBS, K.H., SAUR, W. & SCHULTZ, G. (1976). Reduction of adenylate cyclase activity in lystates of human platelets by the α-adrenergic component of epinephrine. *J. Cyclic Nucleotide Res.*, 2, 381–392.

MARQUIS, N.R., BECKER, J.A. & VIGDAHL, R.L. (1970).
Platelet aggregation, an epinephrine-induced decrease in cyclic-AMP synthesis. *Biochem. Res. Commun.*, 39, 783-789.

MILLS, D.C.B. & ROBERTS, G.C.K. (1967). Effects of adrenaline on human blood platelets. J. Physiol. (Lond.) 193, 443–453.

NEWMAN, K.D., WILLIAMS, L.T., BISHOPRIC, N.H. &

LEFKOWITZ, R.J. (1978). Identification of α-adrenergic receptors in human platelets by [³H]-dihdroergocryptine binding. *J. Clin. Invest.* **61**, 395–402.

O'BRIEN, J.R. (1963). Some effects of adrenaline and antiadrenaline compounds on platelets in vitro and in vivo. Nature, Lond. 200, 763-764.

WILLIAMS, L.T., MULLIKIN, D. & LEFKOWITZ, R.J. (1976). Identification of α-adrenergic receptors in uterine smooth muscle membranes by [³H]-dihydroergocryptine. *J. Biol. Chem.* **251**, 6915–6923.

Binding studies on alpha-adrenoceptors and muscarinic cholinoceptors in rat heart ventricle: effect of chemical sympathectomy

M.S. BRILEY, S.Z. LANGER & D.F. STORY

Synthélabo, L.E.R.S., Department of Biology, 58, rue de la Glacière, 75013 Paris, France

Noradrenaline release in the peripheral nervous system is regulated through a negative feed-back mechanism mediated by presynaptic alpha-adrenoceptors. In addition a number of other presynaptic receptors including inhibitory muscarinic cholinoceptors have been described (Langer, 1977; Starke, 1977).

Rat heart ventricle possesses a rich noradrenergic innervation with mainly postsynaptic beta₁ adrenoceptors. Postsynaptic alpha-adrenoceptors mediating a positive ionotropic effect have also been reported (Wagner & Brodde, 1978).

In the present experiments the binding of two alphaadrenoceptor ligands, [³H]-dihydroergocryptine ([³H]-DHE) and [³H]-WB 4101 (2-([2',6'-dimethoxy] phenoxyethylamine-methylbenzodioxan), and the muscarinic cholinoceptor ligand, [³H]-quinuclidinyl benzilate ([³H]-QNB), to rat heart ventricular membranes were studied in normal animals and animals in which the noradrenergic nerve terminals were destroyed with 6-hydroxydopamine (6-OHDA) pretreatment for two weeks.

The two adrenoceptor ligands, [³H]-DHE and [³H]-WB 4101, each bound with a single high affinity component with apparent dissociation constants (K_D) of 2.3 \pm 3.0 nm and 2.0 \pm 0.6 nm respectively. The maximal binding (Bmax) of [³H]-DHE and [³H]-WB were 374.2 \pm 36.3 fmoles/g tissue and 275.7 \pm 50.5 fmoles/g tissue respectively. Membranes prepared from sympathectomized animals showed decreases in

maximal binding of 59.4% for [³H]-DHE (*P*<0.002) and 24.3% for [³H]-WB 4101 (*P*<0.25) when compared with those prepared from control animals.

The loss of alpha-adrenoceptor binding sites after sympathectomy suggests that some of these sites are located presynaptically on noradrenergic nerve terminals in the rat heart ventricle.

The muscarinic cholinoceptor ligand, [3 H]-QNB, also showed high affinity, single component, binding with a Kd of 0.8 \pm 0.1 nm and Bmax 646.2 \pm 99.2 fmoles/g tissue. After 6-OHDA treatment the maximal binding was not significantly altered (P>0.25).

Our finding of unchanged muscarinic cholinoceptor binding after sympathectomy is in contrast to the recent report of Sharma & Banerjee (1978) who described a significant decrease in [3H]-QNB binding after 6-OHDA treatment. They interpreted their results as a loss of presynaptic cholinoceptors localized in noradrenergic nerve endings. The unchanged (3H]-QNB binding seen in the present experiments suggests however that muscarinic cholinoceptors in the rat heart ventricle are localized mainly postsynaptically.

In summary the present results support a presynaptic location for alpha-adrenoceptors regulating noradrenaline release. Furthermore preliminary experiments have shown that the displacement of the alpha-adrenoceptor ligands from heart ventricle membranes by yohimbine and prazosine is different in normal and 6-OHDA treated animals.

References

LANGER, S.Z. (1977). Presynaptic receptors and their role in the regulation of transmitter release. *Br. J. Pharmac.*, **60**, 481–497.

SHARMA, V.K. & BANERJEE, S.P. (1978). Presynaptic muscarinic cholinergic receptors. *Nature*, **272**, 276–278.

STARKE, K. (1977). Regulation of noradrenaline release by presynaptic receptor systems. *Rev. Physiol. Biochem. Pharmac.*, 77, 1-124.

WAGNER, J. & BRODDE, O.-E. (1978). On the presence and distribution of alpha-adrenoceptors in the heart of various mammalian species. *Naunyn-Schmiedeberg's* Arch. Pharmac., 302, 239-254.